

Tetrahedron 56 (2000) 7691-7703

# Lipid A-Type Pyrancarboxylic Acid Derivatives, their Synthesis and their Biological Activities

Takashi Mochizuki,<sup>a</sup> Yuji Iwano,<sup>b</sup> Masao Shiozaki,<sup>a,\*</sup> Shin-ichi Kurakata,<sup>c</sup> Saori Kanai<sup>c</sup> and Masahiro Nishijima<sup>d</sup>

<sup>a</sup>Exploratory Chemistry Research Laboratories, Sankyo Co. Ltd., Hiromachi 1-2-58, Shinagawa-ku, Tokyo 140-8710, Japan <sup>b</sup>ChemTech Labo, Inc., Hiromachi 1-2-58, Shinagawa-ku, Tokyo 140-8710, Japan

<sup>c</sup>Biological Research Laboratories, Sankyo Co. Ltd., Hiromachi 1-2-58, Shinagawa-ku, Tokyo 140-8710, Japan

<sup>d</sup>Department of Biochemistry and Cell Biology, National Institute of Infectious Diseases, Toyama 1-23-1, Shinjuku-ku,

Tokyo 162-8640, Japan

Received 18 May 2000; accepted 2 August 2000

Abstract—Synthesis of lipid A-type pyrancarboxylic acid derivatives, which have a carboxylic acid group in the anomeric position of the reducing sugar part of the disaccharide instead of the phosphoric acid group in lipid A, is described. We investigated the influence of the substituents in the 2'- and 6'-position of the molecules synthesized on their activities toward human monoblastic U937 cells. It was revealed that a series of compounds, possessing an acetamido group in the 2'-position showed strong LPS-antagonistic activity. © 2000 Elsevier Science Ltd. All rights reserved.

# Introduction

Lipopolysaccharides (LPS)<sup>1</sup> cover the outer surface membrane of Gram-negative bacteria. LPS are known to stimulate the immune system of the host cells, resulting in many pathophysiological events such as fever, depression of blood pressure, platelet aggregation, shock, and organ failure leading to bacterial sepsis.<sup>2</sup> Most of the biological activities of LPS reside in a relatively small portion of the molecule, that is, the terminal disaccharide phospholipid subunit known as lipid A (1, Fig. 1),<sup>1,3,4</sup> which is a hydrophobic anchor substance that links an essentially linear polysaccharide chain to the cell wall. In recent years, a lot of lipid A analogues have been investigated to find LPS antagonists as antisepticemia drugs,<sup>5–7</sup> and also LPS agonists as anticancer drugs. Then, a lot of information on the effect of substitutions in the molecules on the biological activities has been reported. It has been revealed that the phosphoric acid group in the 1-position of lipid A can be exchanged with the carboxymethyl group and dicarboxymethyl group without loss of the biological activities of lipid A.<sup>8,9</sup> In addition, the number of fatty acid chains in lipid A-related compounds appear to be important as to

whether the molecules show LPS-agonistic or -antagonistic activities.  $^{10}$ 

During our investigation of the biological activities of compounds related to GLA-60 (2),<sup>11</sup> a lipid A-related mono-saccharide compound which possesses LPS-agonistic activity, we found that the pyrancarboxylic acid derivative



Figure 1. The structures of *E. coli* lipid A (1), GLA-60 (2), and the pyrancarboxylic acid derivative 3.

*Keywords*: antibacterials; biologically active compounds; glycolipids; lipopolysaccharides.

<sup>\*</sup> Corresponding author. Tel.: +81-3-3492-3131; fax: +81-3-5436-8570; e-mail: shioza@shina.sankyo.co.jp



Figure 2. The structures of lipid A-type pyrancarboxylic acids 4a-c and 5a-c.

**3** showed LPS-antagonistic activity.<sup>12</sup> Then, expecting that disaccharide derivatives like lipid A molecules should have more potent LPS-antagonistic activity, we investigated the synthesis and biological activities of lipid A-type (disaccharide) pyrancarboxylic acid derivatives (Fig. 2).<sup>13</sup> Herein, we disclose the full details of our study of lipid A-type pyrancarboxylic acid derivatives.

We first wanted to examine whether the carboxy group attached directly to the pyran in the anomeric position in the  $\alpha$ -configuration plays a decisive role in LPS-agonism or -antagonism of lipid A-type pyrancarboxylic acid derivatives. If not as such, we also wanted to know whether the phosphate group in the anomeric position of lipid A could be exchanged to a carboxy group, which would be more stable than the former, without loss of biological activities. Further, we tried to investigate the effect of the number of fatty acid chains in the pyrancarboxylic acid derivatives upon the biological activities by changing the substituent in the 2'-position in the molecules from (R)-3-(dodecanoyloxy)tetradecanamido groups (4a-c) to acetamido groups (5a-c). In addition, the effect of a substituent in the 6'-position of the compounds was also examined by comparing the biological activities of 6'-hydroxyl (4a, 5a), 6'-methoxy (4b, 5b), and 6'-fluoro derivatives (4c, 5c).

#### Results

## **Synthesis**

At first, the starting allyl 2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)- $\alpha$ -D-glucopyranoside (6)<sup>3b</sup> was converted to the three different 6-substituted glycosyl donors 9, 13, and 15 as shown in Scheme 1. The 6-hydroxyl group of the diol 6 was selectively protected as a benzyloxycarbonate 7 by treatment with benzylchloroformate and pyridine according to the reported method.<sup>14</sup> After 4-O-phosphorylation of  $\tilde{7}$ with diphenyl chlorophosphate and N,N-dimethylaminopyridine (DMAP), the allyl group in the anomeric position of the thus-obtained 8 was deprotected by treatment with (1,5-cyclooctadiene)bis(methyldiphenylphosphine)iridium(I) hexafluorophosphate in THF and successive hydrolysis with H<sub>2</sub>O-I<sub>2</sub>, affording a 6-O-benzyloxycarbonated intramolecular hemiacetal 9.15 On the other hand, the synthesis of 6-methoxy and 6-fluoro derivatives commenced with selective silvlation of the 6-hydroxyl group of 6 with tertbutyldimethylsilyl chloride and imidazole in DMF to give the 6-O-monosilylated compound 10. Subsequently, 4-Ophosphorylation of 10 with diphenyl chlorophosphate and DMAP, and successive deprotection of the 6-O-silyl group



Scheme 1. Reagents and conditions: (a) CICO<sub>2</sub>Bn, Py, rt, 1 h, 60%; (b) CIPO(OPh)<sub>2</sub>, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 h for 7, 8: 98%, 30 min for 10; (c) [Ir(COD)(P-MePh<sub>2</sub>)<sub>2</sub>]PF<sub>6</sub>, THF, rt, 1 h, then H<sub>2</sub>O–I<sub>2</sub>, 60°C, 1 h, 9: 77%, 13: 73%, 15: 79%; (d) *t*-BuMe<sub>2</sub>SiCl, imidazole, DMF, rt, 1 h, 84%; (e) 3 M HCl aq., THF, 50°C, 1 h, 96% in 2 steps from 10; (f) Me<sub>3</sub>OBF<sub>4</sub>, 2,6-di(*tert*-butyl)-4-methylpyridine, CH<sub>2</sub>Cl<sub>2</sub>, rt, 24 h, 68%; (g) DAST, CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 3 h, 29%.





Scheme 2. Reagents and conditions: (a) PPh<sub>3</sub>, THF, 2 h, then 28% aq. NH<sub>4</sub>OH, 60°C, 3 h; (b) (R)-3-(benzyloxy)tetradecanoic acid, DCC, CH<sub>2</sub>Cl<sub>2</sub>, rt, 15 h, 68% in 2 steps from 16; (c) 4 M HCl, 1,4-dioxane-H<sub>2</sub>O, 60°C, 4 h, then Ph<sub>2</sub>CN<sub>2</sub>, THF, 60°C, 2 h, 33%; (d) Me<sub>2</sub>C(OMe)<sub>2</sub>, p-TsOH, DMF, rt, 3 h, 80%; (e) (R)-3-(benzyloxy)tetradecanoic acid, DCC, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, rt, 16 h, 85%; (f) p-TsOH, MeOH-THF, rt, 3 h, 87%.

with 3 M aqueous hydrochloric acid in THF provided alcohol **11**. After methylation of the 6-hydroxyl group of **11** with trimethyloxonium tetrafluoroborate and 2,6-di(*tert*butyl)-4-methylpyridine, deprotection of the anomeric allyl group of the thus-obtained **12** was accomplished by the same procedure as mentioned above, affording 6-methoxy hemiacetal **13**. Furthermore, after fluorination of the 6-hydroxyl group of **11** with diethylaminosulfur trifluoride (DAST), the anomeric allyl group of the thus-obtained **14** was likewise deprotected to yield 6-fluoro hemiacetal **15**.

Glycosyl acceptor **21** was prepared in accordance with the reported procedure (Scheme 2).<sup>12</sup> The 2-azido group of 2,6-anhydro-3-azido-3-deoxy-5,7-*O*-isopropylidene-D-*glycero*-D-*ido*-heptononitrile (**16**)<sup>12</sup> was transformed to the (*R*)-3-(benzyloxy)tetradecanamido group of **17** by successive treatment with triphenylphosphine and aqueous NH<sub>4</sub>OH, followed by (*R*)-3-(benzyloxy)tetradecanoic acid and dicyclohexyl carbodiimide (DCC). Acidic hydrolysis of the nitrile **17** with 4 M HCl in dioxane–H<sub>2</sub>O, and successive



Scheme 3. Reagents and conditions: (a)  $Cl_3CCN$ , DBU,  $CH_2Cl_2$ , 0°C, 30 min; (b) 21, TMSOTf, MS 4Å,  $CH_2Cl_2$ ,  $-50^{\circ}C$ , 1 h, 22a: 56% in 2 steps from 9, 22b: 57% in 2 steps from 13, 22c: 63% in 2 steps from 15.

esterification of the resulting carboxylic acid with diphenyl diazomethane gave a triol 18.<sup>+</sup> After transformation of 18 to 5,7-*O*-isopropylidene 19 by treatment with 2,2-dimethoxy-propane and a catalytic amount of *p*-toluenesulfonic acid (*p*-TsOH), esterification of the 4-hydroxyl group of 19 with (*R*)-3-(benzyloxy)tetradecanoic acid, DCC, and DMAP yielded compound 20. Then, deprotection of the isopropylidene group of 20 using *p*-TsOH as a catalyst in MeOH-THF furnished glycosyl acceptor 21.

The glycosylation reaction of **9**, **13**, and **15** with **21** was conducted by use of Schmidt's imidate method<sup>16</sup> (Scheme 3). After conversion of the hemiacetals **9**, **13**, and **15** to corresponding trichloroacetimidates with trichloroacetonitrile and a catalytic amount of 1,8-diazabicyclo[5.4.0]-7undecene (DBU), successive glycosylation with **21** by use of trimethylsilyl trifluoromethanesulfonate (TMSOTf) as a catalyst gave rise to only  $\beta$ -oriented pseudodisaccharides **22a**, **22b**, and **22c**, respectively.

Then, the intermediates 22a, 22b, and 22c were converted into two kinds of lipid A-type pyrancarboxylic acid derivatives; ones with a (R)-3-(dodecanoyloxy)tetradecanamido group at the 2'-position in the molecules (Scheme 4) similar to Escherichia coli lipid A, and the others with an acetamido group at the same position (Scheme 5). After deprotection of the trichloroethoxycarbonyl (Troc) group at the 2'-position of 22a-c with zinc dust-acetic acid, we treated the thusobtained amines with (R)-3-(dodecanoyloxy)tetradecanoic acid and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDCI·HCl) to afford **23a**–c.<sup>‡</sup> Hydrogenolysis of this series of compounds using palladium hydroxide on carbon as a catalyst completed the deprotection of the two benzyl ether groups and the diphenylmethyl ester group of 23a-c, and the benzyloxycarbonyl group of 23a to furnish 24a-c. Finally, hydrogenolytic deprotection of each diphenyl phosphate ester group of 24a-c using platinum oxide as a catalyst yielded 4a, 4b, and 4c, respectively

<sup>&</sup>lt;sup> $\dagger$ </sup> This hydrolysis step was thoroughly examined, but gave rise to product **18** in 33% yield at most.

<sup>&</sup>lt;sup>‡</sup> The given yields of **23a** and **23c** are the results of only one experiment each. Higher yields might be obtained with further investigation.

(Scheme 4). We conducted this 2-step reductive deprotection procedure. So we could avoid the troublesome purification of the very polar products at the last step. We excluded the impurities by reduction after the first deprotection step.

On the other hand, after deprotection of the Troc groups of 22a-c by the above-mentioned way, we introduced the acetamido group at the 2'-position with acetic acid and EDCI-HCl as a condensing agent, affording 25a-c. By the same procedure as the synthesis of 4a-c, the two benzyl ether groups, the diphenylmethyl ester group, and the benzyloxycarbonyl group in the case of 25a were deprotected, and then the diphenyl phosphate groups of the thus-obtained 26a-c were deprotected to furnish 5a, 5b, and 5c, respectively (Scheme 5).

## **Biological activity**

The biological activities of the newly synthesized compounds **4a**–**c** and **5a**–**c** were investigated by measuring TNF $\alpha$  production in human monoblastic U937 cells, pretreated with TPA, in the absence (LPS agonism) or presence (LPS antagonism) of LPS (30 ng/ml). As shown in Figs. 3 and 4, compounds **4a**–**c**, which have six fatty acid moieties in each molecule, induced the production of TNF $\alpha$  (LPS-agonistic activity) and showed weak or no inhibitory activity toward LPS-stimulated TNF $\alpha$  production. Compounds **4a** and **4b**, slightly inhibited LPS-induced TNF $\alpha$  production at a higher concentration of 500 nM. On the other hand, compounds **5a**–**c**, which have four lipids, strongly inhibited the production of TNF $\alpha$  induced by LPS



Scheme 4. Reagents and conditions: (a) Zn, AcOH, rt, 3 h; (b) (*R*)-3-(dodecanoyloxy)tetradecanoic acid, EDCI·HCl, CH<sub>2</sub>Cl<sub>2</sub>, rt, 16 h, **23a**: 38% in 2 steps from **22a**, **23b**: 59% in 2 steps from **22b**, **23c**: 24% in 2 steps from **22c**; (c) H<sub>2</sub>, 20% Pd(OH)<sub>2</sub>/C, AcOEt or EtOH, rt, 15–20 h, **24a**: 57%, **24b**: 57%, **24c**: 58%; (d) H<sub>2</sub>, PtO<sub>2</sub>, THF, rt, 15–20 h, **4a**: 88%, **4b**: 49%, **4c**: 73%.



Scheme 5. Reagents and conditions: (a) Zn, AcOH, rt, 5 h; (b) AcOH, EDCI-HCl, CH<sub>2</sub>Cl<sub>2</sub>, rt, 4 h, **25a**: 47% in 2 steps from **22a**, **25b**: 75% in 2 steps from **22b**, **25c**: 57% in 2 steps from **22c**; (c) H<sub>2</sub>, 20% Pd(OH)<sub>2</sub>/C, AcOEt or EtOH, rt, 7–16h, **26a**: 63%, **26b**: 63%, **26c**: 65%; (d) H<sub>2</sub>, PtO<sub>2</sub>, THF, rt, 15–20 h, **5a**: 82%, **5b**: 99%, **5c**: 87%.

in a dose-dependent manner (LPS-antagonistic activity, Fig. 4), with an  $IC_{50}$  value of 7, 10, 9 nM, respectively, but did not show any LPS-agonistic activity (Fig. 3).

## Discussion

The biological activities of the lipid A-type pyrancarboxylic acid derivatives synthesized in this work suggest the following relationships between the structural features of the molecules and their biological activities. Firstly, the LPSagonistic activity of compounds 4a-c, which have a similar acyl substitution pattern to *E. coli* lipid A, and have a carboxylic acid group directly attached to the anomeric position of each molecule instead of the phosphate group in lipid A, revealed that the phosphate group can be replaced with the carboxy group without loss of biological activity. The exchangeability of the phosphate group to the stable carboxy group is useful information for development of lipid A-type molecules, in which instability of the molecules in the anomeric position is often encountered.

Secondly, by comparison of the biological activities of compounds 4a-c with compounds 5a-c, it appears that the number of lipid chains in the molecule is crucial for whether the lipid A-type molecule has LPS-agonistic or -antagonistic activity. Compounds 5a-c, which have four lipid chains in the molecules, have strong LPS-antagonistic activities without any LPS-agonistic activity as lipid IVa<sup>5</sup> (Fig. 5), which also has four lipid chains in the molecule and is known as an LPS-antagonist toward human cells. The importance of the number of lipid chains for biological



**Figure 3.** Production of  $TNF\alpha$  from TPA-treated U937 cells stimulated by lipid A-type pyrancarboxylic acid derivatives. In this experiment, the dosedependency of the response by LPS (data not shown) and *E. coli* lipid A<sup>17</sup> was confirmed. As a control, the amounts of  $TNF\alpha$  produced by TPA-treated U937 cells stimulated with LPS (30 ng/ml) was 1183 pg/ml.



Figure 4. Inhibition of TNF $\alpha$  release from TPA-treated U937 cells stimulated by LPS (30 ng/ml) in the presence of lipid A-type pyrancarboxylic acid derivatives.

activities is obvious. This study provides a new acyl substitution pattern for LPS-antagonistic lipid A-related compounds.

Thirdly, there is no significant difference in LPS-antagonistic activities among the 2'-acetamido derivatives 5a-c. Likewise, compounds 4a-c show almost the same level of LPS-agonistic activity, except for the slight inhibitory activity of a higher concentration of 4c on LPS-induced TNF $\alpha$  production, the reason for which was not elucidated. Therefore, the substituent in the 6'-position of the molecules appears not to be very important for their biological activities. Further studies are necessary to investigate the



slight inhibition of the TNF $\alpha$  production at a higher concentrations of LPS-agonistic compound **4c**.

#### Conclusion

In summary, two kinds of lipid A-type pyrancarboxylic acid derivatives were synthesized. One of them, having four lipid chains in the molecules, showed strong LPS-antagonistic activity. The exchangeability of the phosphate group in the anomeric-position to a stable carboxy group and the importance of the lipid chain number suggest an effective acyl substitution pattern for developing LPS-antagonists.

## **Experimental**

#### General

<sup>1</sup>H NMR spectra were recorded using tetramethylsilane as the internal reference. IR absorption spectra were recorded on a Jasco IR A-2 spectrophotometer, and mass spectra were obtained with a JMS-700 mass spectrometer. 3-Nitrobenzylalcohol was used for a matrix on every measurement.

Figure 5. The structure of lipid IVa (27).

Elemental analyses were performed by the Institute of Science and Technology, Inc. Separation of the compounds by column chromatography was carried out with silica gel 60 (Merck, 230–400 mesh ASTM).

Allyl 6-O-benzyloxycarbonyl-2-deoxy-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycar**bonylamino**)- $\alpha$ -**D**-glucopyranoside (7). To a solution of allyl 2-deoxy-3-O-[(R)-3-tetradecanoyloxytetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)-α-D-glucopyranoside (6,<sup>3b</sup> 438 mg, 0.526 mmol) in THF (30 ml), benzyloxycarbonylchloride (125 mg, 0.735 mmol) and pyridine (62.7 mg, 0.790 mmol) were added. After stirring for 2 h at room temperature, the reaction mixture was concentrated in vacuo, diluted with ethyl acetate, washed with 1 M hydrochloric acid, saturated aqueous NaHCO<sub>3</sub>, and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuo to give a mixture. The mixture was chromatographed on a silica gel column. Elution with hexane-EtOAc (1:1) gave 7 (304 mg, 60% yield) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3472, 1748, 1724, 1540, 1296 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.45–7.29 (5H, m), 5.94–5.79 (1H, m), 5.41–5.05 (7H, m, including 2H, s, at 5.18 ppm), 4.90 (1H, d, J=3.5 Hz), 4.76 (1H, ABq, J=12.0 Hz), 4.65 (1H, AB-q, J=12.0 Hz), 4.45 (2H, d, J=3.3 Hz), 4.20-4.11 (1H, m), 4.01-3.85 (3H, m), 3.66-3.57 (1H, m), 3.40 (1H, d, J=3.9 Hz), 2.56–2.45 (2H, m), 2.29 (2H, t, J=7.5 Hz), 1.68-1.52 (4H, m), 1.30-1.17 (38 H, m), 0.88 (6H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{48}H_{76}NO_{12}Cl_3Na(M+Na)^+$ : 986.4331; found: 986.4332. Anal. Calcd for C<sub>48</sub>H<sub>76</sub>NO<sub>12</sub>Cl<sub>3</sub>+H<sub>2</sub>O: C, 58.62; H, 7.99; N, 1.42; Cl, 10.81. Found: C, 58.67; H, 7.90; N, 1.49; Cl, 11.05.

Allyl 6-O-benzyloxycarbonyl-2-deoxy-4-O-diphenylphosphono-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)-α-D-glucopyranoside (8). To a solution of 7 (278 mg, 0.288 mmol) in  $CH_2Cl_2$ (5 ml), DMAP (70.8 mg, 0.58 mmol) and diphenyl chlorophosphate (155 mg, 0.577 mmol) were added. After stirring for 2 h at room temperature, the reaction mixture was concentrated in vacuo, diluted with ethyl acetate, washed with saturated aqueous NaHCO<sub>3</sub> and brine, and concentrated in vacuo to give a mixture. The mixture was purified using silica gel chromatography. Elution with hexane-EtOAc (5:1) gave 8 (337 mg, 98% yield) as a viscous oil. IR:  $\nu_{\text{max}}$  (neat) 1751, 1267 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.65-7.14 (15H, m), 5.93-5.79 (1H, m), 5.47-5.38 (2H, m), 5.26 (1H, dd, J=28.2, 1.6 Hz), 5.25 (1H, d, J =1.1 Hz), 5.13-5.01 (3H, m, including 2H, AB-q, J=12.1 Hz, at 5.11, 5.01 ppm), 4.79 (1H, AB-q, J=12.0 Hz), 4.74 (1H, q, J=9.4 Hz), 4.60 (1H, AB-q, J=12.0 Hz), 4.35-4.11 (3H, m), 4.08-3.95 (3H, m), 2.45 (2H, d, J=6.3 Hz), 2.14 (2H, t, J=7.6 Hz), 1.56–1.40 (4H, m), 1.33–0.92 (38H, m), 0.88 (6H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for C<sub>60</sub>H<sub>85</sub>NO<sub>15</sub>Cl<sub>3</sub>PK (M+K)<sup>+</sup>: 1234.4359; found: 1234.4381. Anal. Calcd for C<sub>60</sub>H<sub>85</sub>NO<sub>15</sub>Cl<sub>3</sub>P: C, 60.17; H, 7.15; N, 1.17; Cl, 8.88; P, 2.59. Found: C, 60.29; H, 6.95; N, 1.30; Cl, 8.68; P, 2.57.

**6-O-Benzyloxycarbonyl-2-deoxy-4-O-diphenylphosphono-3-O-**[(*R*)-**3-(tetradecanoyloxy)tetradecanoyl]-2-**(**2,2,2-trichloroethoxycarbonylamino)-D-glucopyranose** (**9).** A solution of **8** (881 mg, 0.735 mmol) and (1,5-cyclo-

octadiene)bis(diphenylmethylphosphine)iridium(I) hexafluorophosphate (33.9 mg, 0.0401 mmol) in THF (5 ml) was replaced with a hydrogen atmosphere. Stirring for 1 min, the color of the solution turned colorless from redbrown, then immediately the reaction mixture was replaced with a nitrogen atmosphere. After stirring for 1 h at room temperature, H<sub>2</sub>O (0.5 ml) and I<sub>2</sub> (280 mg, 1.10 mmol) were added to the mixture, and the mixture was stirred for 1 h at 60°C. The reaction mixture was quenched with aqueous 5% Na<sub>2</sub>SO<sub>3</sub>, extracted with EtOAc, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuo to give a mixture. The mixture was purified by silica gel chromatography. Elution with hexane-EtOAc (4:1) gave 9 (656 mg, 77% yield) as a powder. IR:  $\nu_{max}$  (KBr) 3344 (broad), 1749, 1490, 1269 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.65–7.11 (15H, m), 5.58–5.45 (2H, m), 5.32 (1H, d, J=3.4 Hz), 5.13– 5.01 (3H, m, including 2H, AB-q, J=12.0 Hz, at 5.11, 5.03 ppm), 4.73 (1H, q, J=9.2 Hz), 4.64 (1H, AB-q, J=12.0 Hz), 4.52 (1H, AB-q, J=12.0 Hz), 4.37–4.18 (3H, m), 4.03–3.94 (1H, m), 2.45 (2H, d, J=6.3 Hz), 2.16 (2H, t, J=7.6 Hz), 1.67-1.33 (4H, m), 1.30-1.11 (38H, m), 0.88 (6H, t, J=6.6 Hz). Anal. Calcd for  $C_{57}H_{81}NO_{15}Cl_3P+H_2O$ : C, 58.24; H, 7.12; N, 1.19; Cl, 9.05; P, 2.63. Found: C, 58.48; H, 7.04; N, 1.23; Cl, 9.32; P, 2.66.

Allyl 6-O-tert-butyldimethylsilyl-2-deoxy-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)-α-D-glucopyranoside (10). To a solution of 6 (2.37 g, 2.86 mmol) in DMF (30 ml), imidazole (398 mg, 5.85 mmol) and tert-butyldimethylsilyl chloride (470 mg, 3.12 mmol) were added. After stirring for 1 h at room temperature, the reaction mixture was diluted with diethyl ether, washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuo to give a mixture. The mixture was chromatographed on a silica gel column. Elution with hexane-EtOAc (10:1) gave 10 (2.27 g, 84% yield) as a viscous oil. IR:  $\nu_{\text{max}}$  (neat) 2926, 2856, 1743 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 5.94–5.84 (1H, m), 5.44 (1H, d, J=9.5 Hz, NH), 5.32–5.12 (4H, m), 4.89 (1H, d, J=3.6 Hz), 4.74, 4.68 (2H, AB-q, J=12.5 Hz), 4.19 (1H, dd, J=12.5, 5.1 Hz), 3.99 (1H, dd, J=12.5, 6.6 Hz), 3.94-3.84 (2H, m), 3.73-3.63 (3H, m), 3.38 (1H, broad s, H), 2.59 (1H, dd, J=15.0, 7.7 Hz), 2.50 (1H, dd, J=15.4, 4.4 Hz), 2.28 (2H, t, J=7.7 Hz), 1.64-1.58 (2H, m), 1.36-1.20 (40H, m), 0.96-0.86 (15H, m), 0.10 (3H, s), 0.07 (3H, s). High Resolution MS (FAB, positive), calcd for C46H84NO10Cl3SiNa  $(M+Na)^+$ : 966.4828; found: 966.4811.

Allyl 2-deoxy-4-*O*-diphenylphosphono-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)- $\alpha$ -D-glucopyranoside (11). To a solution of 10 (2.27 g, 2.40 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 ml), DMAP (589 mg, 4.82 mmol) and diphenyl chlorophosphate (1.30 g, 4.84 mmol) were added. After stirring for 30 min at room temperature, the reaction mixture was quenched with water, extracted with EtOAc, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuo to give a mixture, which was further used without purification. To a solution of the mixture above obtained in THF (30 ml), aqueous 3 M hydrochloric acid was added. After stirring for 1 h at 50°C, the reaction mixture was concentrated in vacuo to give a crude product. The crude product was

7697

purified by silica gel chromatography. Elution with hexane– EtOAc (2:1) gave **11** (2.46 g, 96% yield) as a viscous oil. IR:  $\nu_{max}$  (neat) 3439, 2926, 2855, 1745 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 7.47–7.12 (10H, m), 5.92–5.83 (1H, m), 5.47 (1H, t, *J*=9.6 Hz), 5.39 (1H, d, *J*=9.7 Hz), 5.31 (1H, dd, *J*=17.3, 1.3 Hz), 5.27–5.23 (1H, m), 5.13 (1H, t, *J*=6.3 Hz), 4.97 (1H, d, *J*=3.5 Hz), 4.80–4.73(1H, m), 4.76 (1H, AB-q, *J*=12.0 Hz), 4.65 (1H, AB-q, *J*=12.0 Hz), 4.21–4.16 (1H, m), 4.08–3.98 (2H, m), 3.75 (1H, d, *J*=9.8 Hz), 3.64–3.62 (2H, m), 3.32–3.28 (1H, m), 2.42– 2.40 (2H, m), 2.18 (2H, t, *J*=7.6 Hz), 1.60–1.40 (2H, m), 1.38–1.16 (40H, m), 0.88 (3H, t, *J*=6.8 Hz). High Resolution MS (FAB, positive), calcd for C<sub>52</sub>H<sub>79</sub>NO<sub>13</sub>Cl<sub>3</sub>PNa (M+Na)<sup>+</sup>: 1084.4252; found: 1084.4258.

Allyl 2-deoxy-4-O-diphenylphosphono-6-O-methyl-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)- $\alpha$ -D-glucopyranoside (12). To a solution of 11 (13.1 mg, 0.012 mmol) in  $CH_2Cl_2$ (1 ml), a solution of trimethyloxonium tetrafluoroborate (36.0 mg, 0.243 mmol) and 2,6-di(tert-butyl)-4-methylpyridine (9.1 mg, 0.044 mmol) were added. After stirring for 24 h at room temperature, the reaction mixture was quenched with water, extracted with EtOAc, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuo to give a mixture. The mixture was chromatographed on a silica gel column. Elution with hexane-EtOAc (5:1) gave 12 (9.0 mg, 68% yield) as a viscous oil. IR:  $\nu_{max}$  (KBr) 1748, 1516 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.37–7.17 (10H, m), 5.96-5.81 (1H, m), 5.48-5.22 (4H, m), 5.16-5.00 (1H, m), 4.96 (1H, d, J=3.5 Hz), 4.84-4.73 (2H, m, including 1H, AB-q, J=11.8 Hz at 4.78 ppm), 4.61 (1H, AB-q, J=11.8 Hz), 4.25-4.18 (1H, m), 4.05-3.90 (3H, m), 3.52-3.42 (2H, m), 3.20 (3H, s), 2.45 (2H, d, J=6.3 Hz), 2.13 (2H, t, J=7.5 Hz), 1.61-1.13 (42 H, m), 0.88 (6H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{53}H_{81}NO_{13}Cl_3PNa (M+Na)^+$ : 1098.4409; found: 1098.4390. Anal. Calcd for C<sub>53</sub>H<sub>81</sub>NO<sub>13</sub>Cl<sub>3</sub>P: C, 59.08; H, 7.58; N, 1.30; Cl, 9.87; P, 2.87. Found: C, 58.40; H, 7.71; N, 1.26; Cl, 10.15; P, 2.84.

**2-Deoxy-4-***O***-diphenylphosphono-6***-O***-methyl-3***-O*-[(*R*)-**3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)-D-glucopyranose (13).** Compound **12** (898 mg, 0.833 mmol) was treated as described in the formation of **9** from **8** to give **13** (629 mg, 73% yield) as a powder. IR:  $\nu_{max}$  (KBr) 3399 (broad), 1728, 1525 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.37–7.18 (10H, m), 5.66 (1H, d, *J*=9.5 Hz), 5.50 (1H, dd, *J*=10.3, 9.8 Hz), 5.32 (1H, d, *J*=3.1 Hz), 5.15–5.06 (1H, m), 4.79–4.62 (3H, m), 4.32 (1H, broad s, OH), 4.22–4.18 (1H, m), 3.98 (1H, dt, *J*<sub>d</sub>=3.1 Hz, *J*<sub>t</sub>=10.1 Hz), 3.52–3.37 (2H, m), 3.20 (3H, s), 2.44 (2H, d, *J*=6.1 Hz), 2.15 (2H, t, *J*=7.6 Hz), 1.55–1.11 (42 H, m), 0.88 (6H, t, *J*=6.6 Hz). High Resolution MS (FAB, positive), calcd for C<sub>50</sub>H<sub>78</sub>NO<sub>13</sub>Cl<sub>3</sub>P (M+H)<sup>+</sup>: 1036.4295; found: 1036.4297.

Allyl 2,6-deoxy-4-*O*-diphenylphosphono-6-fluoro-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)- $\alpha$ -D-glucopyranoside (14). To a solution of 11 (2.38 g, 2.24 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (100 ml), diethylaminosulfurtrifluoride (2 ml, 15.1 mmol) was added at  $-35^{\circ}$ C. After stirring for 3 h at 0°C, the reaction mixture was quenched with aqueous phosphate buffer (pH 7), extracted with EtOAc, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuo to give a mixture. The mixture was chromatographed on a silica gel column. Elution with hexane-EtOAc (2:1) gave 14 (683 mg, 29% yield) as a viscous oil. IR:  $\nu_{\rm max}$  (KBr) 1747, 1591, 1516, 1491 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 7.36-7.16 (10H, m), 5.91-5.84 (1H, m), 5.48-5.40 (2H, m), 5.34-5.24 (2H, m), 5.13-5.10 (1H, m), 4.98 (1H, d, J=3.7 Hz), 4.80 (1H, AB-q, J=12.5 Hz), 4.72 (1H, q, J=9.5 Hz), 4.58 (1H, AB-q, J=12.5 Hz), 4.46 (2H, dd, J=47.2, 2.6 Hz), 4.23-4.18 (1H, m), 4.06-3.95 (3H, m), 2.45 (2H, d, J=6.6 Hz), 2.14 (2H, t, J=7.3 Hz), 1.60-1.17 (44H, m), 0.88 (6H, t, J=7.0 Hz). High Resolution MS (FAB, positive), calcd for  $C_{52}H_{78}NO_{12}Cl_3FPNa$  (M+Na)<sup>+</sup>: 1086.4209; 1086.4207. Anal. Calcd found: for C<sub>52</sub>H<sub>78</sub>NO<sub>12</sub>Cl<sub>3</sub>FP: C, 58.62; H, 7.38; N, 1.32; F, 1.78; Cl, 9.98; P, 2.91. Found: C, 58.65; H, 7.41; N, 1.24; F, 1.67; Cl, 9.89; P. 3.04.

2,6-Deoxy-4-O-diphenylphosphono-6-fluoro-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)-D-glucopyranose (15). Compound 14 (683.4 mg, 0.641 mmol) was treated as described in the formation of 9 from 8 to give 15 (518 mg, 79% yield) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3424 (broad), 1728, 1524 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.36-7.17 (10H, m), 5.66 (1H, d, J=9.5 Hz), 5.55 (1H, t, J=9.9 Hz), 5.37 (1H, d, J=3.7 Hz), 5.13-5.09 (1H, m), 4.75 (1H, AB-q, J=11.7 Hz), 4.72 (1H, q, J=9.5 Hz), 4.64 (1H, AB-q, J=11.7 Hz), 4.46 (2H, d, J=47.6 Hz), 4.26 (1H, dd, J=12.5, 6.2 Hz), 4.03-3.97 (1H, m), 2.45 (2H, d, J=6.6 Hz), 2.15 (2H, t, J=7.3 Hz), 1.53-1.20 (42 H, m), 0.88 (6H, t, J=7.0 Hz). High Resolution MS (FAB, positive), calcd for C49H75NO12Cl3FP (M+H)<sup>+</sup>: 1024.4076; found: 1024.4027. Anal. Calcd for C<sub>49</sub>H<sub>74</sub>NO<sub>12</sub>Cl<sub>3</sub>FP: C, 57.39; H, 7.27; N, 1.37; F, 1.85; Cl, 10.37; P, 3.02. Found: C, 57.11; H, 7.05; N, 1.42; F, 1.96; Cl, 10.74; P, 2.90.

2.6-Anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-3deoxy-5,7-O-isopropylidene-D-glycero-D-ido-heptononitrile (17). To a solution of 2,6-anhydro-3-azido-3-deoxy-5,7-O-isopropylidene-D-glycero-D-ido-heptononitrile (16,<sup>12</sup> 10.1 g, 39.6 mmol) in THF (200 ml), triphenylphosphine (11.4 g, 43.6 mmol) was added at room temperature. After stirring for 2 h, 28% aqueous NH<sub>4</sub>OH was added to the reaction mixture, which was further stirred at 60°C for 3 h. Then, the reaction mixture was concentrated in vacuo, and coevaporated with toluene to remove H<sub>2</sub>O. The obtained precipitate was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (100 ml), and to the solution, a solution of (R)-3-(benzyloxy)tetradecanoic acid (13.3 g, 39.8 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (70 ml) and a solution of DCC (8.25 g, 40.0 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 ml) were added. After stirring for 15 h, the resulting white precipitate was removed by filtration, and the solution was concentrated in vacuo to give a mixture. The mixture was purified using silica gel chromatography. Elution with hexane-EtOAc (2:1) gave 17 (14.6 g, 68% yield) as an amorphous film. IR:  $\nu_{\text{max}}$  (neat) 3418, 1644, 1628 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.40–7.32 (10H, m), 7.03 (1H, d, J=6.0 Hz), 5.20 (1H, d, J=6.0 Hz), 4.64 (1H, ABq, J=11.2 Hz), 4.54 (1H, AB-q, J=11.2 Hz), 4.06 (1H, m), 3.91 (1H, dd, J=9.7, 4.3 Hz), 3.82 (1H, m), 3.72 (1H, t,

J=10.4-9.8 Hz), 3.65 (1H, m), 3.55–3.44 (2H, m), 2.58– 2.41 (2H, m), 1.74–1.50 (2H, m), 1.49 (3H, s), 1.43 (3H, s), 1.26 (18H, m), 0.88 (3H, t, J=7.0-6.2 Hz). High Resolution MS (FAB, positive), calcd for  $C_{31}H_{49}N_2O_6$  (M+H)<sup>+</sup>: 545.3591; found: 545.3579.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-3-deoxy-D-glycero-D-ido-heptonate (18). To a solution of nitrile 17 (3.44 g, 3.99 mmol) in dioxane (22 ml)-H<sub>2</sub>O (2.2 ml), 4 M hydrogen chloride dioxane solution was added. After stirring for 4 h at 60°C, the reaction mixture was concentrated in vacuo, giving a precipitate. The precipitate was dissolved in dimethylformamide (DMF, 16 ml), treated with diphenyldiazomethane (3.10 g, 15.9 mmol), and stirred for 2 h at 60°C. The reaction mixture was concentrated in vacuo to give a mixture. The mixture was purified using silica gel chromatography. Elution with EtOAc, and then EtOAc-MeOH (10:1) gave 18 (897 mg, 33% yield) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3324, 1734, 1645, 1538 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.41-7.21 (15H, m), 6.98 (1H, d, J=9.1 Hz), 6.82 (1H, s), 4.55 (1H, d, J=5.7 Hz), 4.40 (2H, AB-q, J=11.5 Hz), 4.33 (1H, m), 3.81-3.57 (6H, m), 3.28 (1H, m), 2.87 (1H, broad s), 2.42-2.27 (2H, m), 1.65-1.40 (2H, m), 1.30-1.25 (18H, m), 0.88 (3H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{41}H_{56}NO_8 (M+H)^+$ : 690.4006; found: 690.4016. Anal. Calcd for C<sub>41</sub>H<sub>55</sub>NO<sub>8</sub>: C, 71.37; H, 8.04; N, 2.03. Found: C, 71.25; H, 8.17; N, 2.00.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-3-deoxy-5,7-O-isopropylidene-D-glycero-Dido-heptonate (19). A solution of triol 18 (781 mg, 1.13 mmol) and p-TsOH (52 mg) in DMF (8 ml) and 2,2dimethoxypropane (8 ml) was stirred for 3 h at room temperature, diluted with diethyl ether, extracted with 10% aqueous NaHCO<sub>3</sub>, washed with brine, and dried over MgSO<sub>4</sub>. The mixture was filtered, concentrated in vacuo, and purified with silica gel chromatography. Elution with hexane-EtOAc (2:1) gave 19 (660 mg, 80% yield) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3376, 1728, 1640 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.38-7.24 (15H, m), 6.84 (1H, s), 6.83 (1H, d, J=6.9 Hz), 4.56 (1H, d, J=5.9 Hz), 4.51-4.35 (3H, m, including 1H, d, J=5.9 Hz, at 4.41 ppm), 3.86-3.62 (4H, m), 3.56 (1H, t, J=9.3 Hz), 3.26-3.17 (1H, m), 2.54 (1H, d, J=2.8 Hz), 2.44-2.29 (2H, m), 1.65-1.36 (8H, m, containing two 3H, s, at 1.48 and 1.37 ppm), 1.29-1.23 (18H, m), 0.88 (3H, t, J=6.8-6.4 Hz). High Resolution MS (FAB, positive), calcd for  $C_{44}H_{60}NO_8 (M+H)^+$ : 730.4319; found: 730.4307. Anal. Calcd for C44H59NO8: C, 72.40; H, 8.15; N, 1.92. Found: C, 71.95; H, 7.85; N, 2.22.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3deoxy-5,7-O-isopropylidene-D-glycero-D-ido-heptonate (20). To a solution of alcohol 19 (610 mg, 0.835 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (8 ml), (R)-3-(benzyloxy)tetradecanoic acid (317 mg, 0.947 mmol), DCC (207 mg, 1.00 mmol), and DMAP (124 mg, 1.01 mmol) were added at 24°C. After stirring for 16 h at 24°C, the reaction mixture was diluted with ethyl acetate, washed with 0.1 M aqueous HCl, saturated aqueous NaHCO<sub>3</sub>, and brine in order, and dried over MgSO<sub>4</sub>. The mixture was filtered, concentrated in vacuo, and purified using silica gel chromatography. Elution with hexane–EtOAc (10:1) gave **20** (747 mg, 85% yield) as a powder. IR:  $\nu_{max}$  (KBr) 2920, 1740 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.54–7.08 (20H, m), 6.85 (1H, s), 6.56 (1H, d, *J*=8.8 Hz), 5.40 (1H, t, *J*=9.8 Hz), 4.60–4.37 (6H, m), 3.81–3.63 (5H, m), 3.31–3.22 (1H, m), 2.65 (1H, dd, *J*=15.1, 6.4 Hz), 2.39 (1H, dd, *J*=15.3, 5.9 Hz), 2.20–2.04 (2H, m), 1.66–1.15 (46H, m), 0.88 (6H, t, *J*=6.8–6.4 Hz). High Resolution MS (FAB, positive), calcd for C<sub>65</sub>H<sub>91</sub>NO<sub>10</sub>Na (M+Na)<sup>+</sup>: 1068.6541; found: 1068.6543. Anal. Calcd for C<sub>65</sub>H<sub>91</sub>NO<sub>10</sub>: C, 74.61; H, 8.77; N, 1.34. Found: C, 74.89; H, 8.68; N, 1.28.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3deoxy-D-glycero-D-ido-heptonate (21). To a solution of isopropylidene 20 (1.09 g, 1.04 mmol) in THF (15 ml) and MeOH (15 ml), p-TsOH (235 mg) was added at 24°C. After stirring for 3 h at 24°C, the reaction mixture was concentrated in vacuo, and purified with silica gel chromatography. Elution with hexane–EtOAc (2:1) gave 21 (906 mg, 87%) yield) as a powder. IR:  $\nu_{max}$  (KBr) 3350, 1733, 1649, 1536 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.33–7.08 (20H, m), 6.85 (1H, s), 6.61 (1H, d, J=8.8 Hz), 5.27 (1H, t, J=9.9 Hz), 4.63 (1H, d, J=5.8 Hz), 4.50-4.32 (5H, m, including two 2H, s, at 4.48 and 4.37 ppm), 3.99-3.80 (1H, m), 3.73-3.58 (5H, m), 3.37-3.30 (1H, m), 3.14-2.76 (1H, broad s), 2.62-2.42 (2H, m), 2.22-2.14 (2H, m), 1.69-1.14 (40H, m), 0.88 (6H, t, J=6.4 Hz). High Resolution MS (FAB, positive), calcd for  $C_{62}H_{88}NO_{10} (M+H)^+$ : 1006.6408; found: 1006.6397. Anal. Calcd for C<sub>62</sub>H<sub>87</sub>NO<sub>10</sub>: C, 74.00; H, 8.71; N, 1.39. Found: C, 74.19; H, 8.67; N, 1.40.

Diphenylmethyl 2,6-anhydro-7-O-[6-O-benzyloxycarbonyl-2-deoxy-4-O-diphenylphosphono-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2-trichloroethoxycarbonylamino)- $\beta$ -D-glucopyranosyl]-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3-deoxy-D-glycero-D-ido-heptonate (22a). To a solution of 9 (249 mg, 0.215 mmol) and trichloroacetonitrile (200 mg, 2.00 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (3.0 ml), a solution of DBU (6.8 mg, 0.0446 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1.5 ml) was added at 0°C under nitrogen. After stirring for 30 min at 0°C, the reaction mixture was concentrated in vacuo, and rapidly chromatographed with hexane-EtOAc (2:1). The eluent was concentrated in vacuo to give a crude imidate, which was immediately used for the next reaction without further purification. To a suspension in CH<sub>2</sub>Cl<sub>2</sub> (4.0 ml) of the thus-obtained imidate, diol 21 (217 mg, 0.216 mmol) and molecular sieves 4Å dried in advance, TMSOTf (4.8 mg, 0.0217 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (50 ml) was added at  $-50^{\circ}$ C. After stirring for 1 h at  $-50^{\circ}$ C, the mixture was quenched with saturated NaHCO<sub>3</sub> aqueous solution, extracted with EtOAc, and the extract was washed with brine, dried over Na2SO4, filtered, and concentrated in vacuo to give a mixture. The mixture was purified with silica gel chromatography. Elution with hexane-EtOAc (3:1) gave 22a (289 mg, 56% yield) as a powder. IR:  $\nu_{\text{max}}$ (KBr) 3343, 1735, 1531, 1268 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.38–7.11 (35H, m), 6.84 (1H, s), 6.51 (1H, d, J=8.9 Hz), 5.71 (1H, d, J=6.8 Hz), 5.55–5.48 (1H, m), 5.32-5.01 (4H, m, including 1H, AB-q, J=12.3 Hz at 5.03 ppm and 1H, AB-q, J=12.3 Hz at 5.12 ppm), 4.99 (1H, d, J=9.7 Hz), 4.75–4.61 (4H, m), 4.57–4.29 (6H, m, including 1H, s at 4.33 ppm, 1H, s, at 4.35 ppm, 1H, s at 4.48 ppm, and 1H, s at 4.49 ppm), 4.22–4.16 (1H, m), 4.01–3.95 (1H, m), 3.85–3.80 (1H, m), 3.74–3.54 (4H, m), 3.45–3.34 (2H, m), 2.82 (1H, broad s), 2.64–2.13 (8H, m), 1.67–1.11 (82H, m), 0.88 (12H, t, J=6.5 Hz). High Resolution MS (FAB, positive), calcd for C<sub>119</sub>H<sub>166</sub>N<sub>2</sub>O<sub>24</sub>Cl<sub>3</sub>PNa (M+Na)<sup>+</sup>: 2166.0531; found: 2166.0549. Anal. Calcd for C<sub>119</sub>H<sub>166</sub>N<sub>2</sub>O<sub>24</sub>Cl<sub>3</sub>P: C, 66.61; H, 7.80; N, 1.31; Cl, 4.96; P, 1.44. Found: C, 66.14; H, 7.59; N, 1.29; Cl, 5.16; P, 1.71.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3deoxy-7-0-[2-deoxy-4-0-diphenylphosphono-6-0-methyl-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2trichloroethoxycarbonylamino)-B-D-glucopyranosyl]-Dglycero-D-ido-heptonate (22b). Compound 13 (21.5 mg, 0.021 mmol) was treated as described in the formation of **22a** from **9** to give **22b** (23.3 mg, 57% yield) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3550–3360, 1738, 1531 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.36–7.15 (30H, m), 6.84 (1H, s), 6.50 (1H, d, J=8.8 Hz), 5.62 (1H, d, J=6.6 Hz), 5.47 (1H, t, J=9.8 Hz), 5.31–5.17 (2H, m), 4.92 (1H, d, J=8.3 Hz), 4.75-4.61 (4H, m), 4.50-4.34 (5H, m), 4.01-3.97 (1H, m), 3.86-3.73 (2H, m), 3.65-3.47 (3H, m), 3.44-3.38 (4H, m), 3.20 (3H, s), 2.65-2.14 (8H, m), 1.66-1.11 (94H, m), 0.88 (12H, t, J=6.1-6.9 Hz). High Resolution MS (FAB, positive), calcd for C<sub>112</sub>H<sub>162</sub>N<sub>2</sub>O<sub>22</sub>Cl<sub>3</sub>PNa (M+Na)<sup>+</sup>: 2046.0320; found: 2046.0288. Anal. Calcd for C<sub>112</sub>H<sub>162</sub>N<sub>2</sub>O<sub>22</sub>Cl<sub>3</sub>P: C, 66.40; H, 8.06; N, 1.38; Cl, 5.25; P, 1.53. Found: C, 66.32; H, 7.85; N, 1.37; Cl, 5.41; P, 1.76.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3deoxy-7-O-[2,6-deoxy-4-O-diphenylphosphono-6-fluoro-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-2-(2,2,2trichloroethoxycarbonylamino)-β-D-glucopyranosyl]-Dglycero-D-ido-heptonate (22c). Compound 15 (312 mg, 0.304 mmol) was treated as described in the formation of **22a** from **9** to give **22c** (385 mg, 63% yield) as a powder. IR:  $\nu_{\rm max}$  (KBr) 3424, 1732, 1654, 1591, 1491, 1189 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 7.35-7.14 (30H, m), 6.84 (1H, s), 6.52 (1H, d, J=8.8 Hz), 5.75-5.69 (1H, m), 5.58-5.50 (1H, m), 5.29-5.16 (2H, m), 5.02 (1H, d, J=8.1 Hz), 4.77-4.70 (2H, m), 4.66–4.57 (3H, m), 4.52–4.38 (5H, m), 4.34 (1H, d, J=5.1 Hz), 4.01 (1H, d, J=7.4 Hz), 3.84-3.81 (1H, m), 3.73-3.59 (4H, m), 3.48-3.43 (1H, m), 3.39-3.32 (1H, m), 2.77 (1H, broad s), 2.60 (1H, dd, J=8.1, 15.4 Hz), 2.45 (1H, d, J=5.1 Hz), 2.41 (1H, d, J=4.4 Hz), 2.32 (1H, dd, J=8.1, 15.4 Hz), 2.24-2.13 (4H, m), 1.30-1.08 (82H, m), 0.88 (12H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{111}H_{159}N_2O_{21}Cl_3FPNa$  (M+Na)<sup>+</sup>: 2034.0120; found: 2034.0135. Anal. Calcd for C<sub>111</sub>H<sub>159</sub>N<sub>2</sub>O<sub>21</sub>Cl<sub>3</sub>FP: C, 66.20; H, 7.96; N, 1.39; Cl, 5.28; F, 0.94; P, 1.54. Found: C, 65.81; H, 7.69; N, 1.49; Cl, 5.32; F, 0.97; P, 1.29.

Diphenylmethyl 2,6-anhydro-7-O-[6-O-benzyloxycarbonyl-2-deoxy-4-O-diphenylphosphono-2-[(R)-3-(dodecanoyloxy)tetradecanamido]-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]- $\beta$ -D-glucopyranosyl]-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3-deoxy-D-glycero-D-ido-heptonate (23a). To a solution of 22a (174 mg, 0.080 mmol) in acetic acid (5.0 ml), zinc dust (745 mg, 11.4 mmol) was added. After vigorously stirring for 3 h at room temperature, the solution was filtered through Celite to remove the zinc dust. The filtrate was concentrated in vacuo to give a residue, which was dissolved in CHCl<sub>3</sub>, washed with saturated aqueous NaHCO<sub>3</sub> solution, dried with MgSO<sub>4</sub>, and filtered. The filtrate was concentrated in vacuo, and dissolved in  $CH_2Cl_2$  (2.0 ml). To this solution, were added (*R*)-3-(dodecanoyloxy)tetradecanoic acid (34.7 mg, 0.081 mmol) and EDCI·HCl (31.0 mg, 0.162 mmol) at room temperature, and the mixture was allowed to stand for 16 h at room temperature. Then, the reaction mixture was concentrated in vacuo, and chromatographed on a silica gel column eluted with hexane-EtOAc (3:1) to give 23a (71.8 mg, 38%) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3341, 1732, 1651, 1493 cm<sup>-1</sup>. <sup>г</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.58–7.10 (35H, m), 6.83 (1H, s), 6.50 (1H, d, J=8.8 Hz), 6.42 (1H, d, J=7.0 Hz), 5.51–5.47 (1H, m), 5.32–5.28 (1H, m), 5.18– 4.99 (5H, m, containing 1H, AB-q, *J*=12.2 Hz at 5.11 ppm, 1H, AB-q, J=12.2 Hz at 5.01 ppm), 4.64–4.29 (9H, m), 4.24-4.17 (1H, m), 4.10-3.97 (1H, m), 3.83-3.60 (5H, m), 3.43-3.39 (1H, m), 2.61-2.08 (12H, m), 1.66-1.14 (120H, m), 0.88 (18H, t, J=6.2 Hz). High Resolution MS (FAB, positive), calcd for  $C_{142}H_{213}N_2O_{25}PNa (M+Na)^+$ : 2400.5093; found: 2400.5078. Anal. Calcd for C<sub>142</sub>H<sub>213</sub>N<sub>2</sub>O<sub>25</sub>P+H<sub>2</sub>O: C, 71.15; H, 9.04; N, 1.17; P, 1.29. Found: C, 71.10; H, 8.78; N, 1.13; P, 1.04.

Diphenylmethyl 2,6-anhydro-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3deoxy-7-O-[2-deoxy-4-O-diphenylphosphono-2-[(R)-3-(dodecanovloxy)tetradecanamido]-6-O-methyl-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-B-D-glucopyranosyl]-D-glycero-D-ido-heptonate (23b). Compound 22b (189 mg, 0.093 mmol) was treated as described in the formation of **23a** from **22a** to give **23b** (123 mg, 59%) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 1736, 1659 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.42-7.14 (30H, m), 6.83 (1H, s), 6.50 (1H, d, J=9.3 Hz), 6.35 (1H, d, J=7.3 Hz), 5.43 (1H, t, J=9.3 Hz), 5.31 (1H, dd, J=10.5, 9.2 Hz), 5.17-5.12 (2H, m), 5.04 (1H, d, J=8.6 Hz), 4.68-4.34 (6H, m), 3.98-3.40 (11H, m), 3.22-3.18 (1H, broad s), 3.18 (3H, s), 2.68-2.04 (12H, m), 1.74-1.23 (120H, m), 0.88 (18H, t, J=6.3 Hz). High (FAB, calcd Resolution MS positive), for  $(M+Na)^{+}$ : found: C135H209N2O23PNa 2280.4882; 2280.4871. Anal. Calcd for C135H209N2O23P+H2O: C, 71.21; H, 9.34; N, 1.23; P, 1.36. Found: C, 71.21; H, 9.10; N, 1.28; P, 1.13.

Diphenylmethyl 2,6-anhydro-3-[(*R*)-3-(benzyloxy)tetradecanamido]-4-*O*-[(*R*)-3-(benzyloxy)tetradecanoyl]-3deoxy-7-*O*-[2,6-deoxy-4-*O*-diphenylphosphono-2-[(*R*)-3-(dodecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-D-glycero-D-ido-heptonate (23c). Compound 22c (123 mg, 0.061 mmol) was treated as described in the formation of 23a from 22a to give 23c (33.1 mg, 24%) as a powder. IR:  $\nu_{max}$  (KBr) 1737, 1666, 1193 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 7.33-7.13 (30H, m), 6.84 (1H, s), 6.51 (1H, d, *J*=8.8 Hz), 6.44 (1H, d, *J*=6.6 Hz), 5.54 (1H, t, *J*= 9.5 Hz), 5.31 (1H, t, *J*=9.5 Hz), 5.20 (1H, d, *J*=8.1 Hz), 5.19-5.13 (2H, m), 4.64 (1H, d, *J*=5.1 Hz), 4.62-4.31 (7H, m), 4.02 (1H, d, J=9.5 Hz), 3.89–3.81 (1H, m), 3.78– 3.60 (5H, m), 3.48–3.44 (2H, m), 2.62 (1H, q, J=7.7 Hz), 2.47–2.10 (11H, m), 1.81–1.25 (120H, m), 0.88 (18H, t, J=7.3-5.1 Hz). High Resolution MS (FAB, positive), calcd for  $C_{134}H_{206}N_2O_{22}FPNa$  (M+Na)<sup>+</sup>: 2268.4682. Found: 2268.4746.

2,6-Anhydro-3-deoxy-7-O-[2-deoxy-4-O-diphenylphosphono-2-[(R)-3-(dodecanoyloxy)tetradecanamido]-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (24a). To a solution of 23a (73.4 mg, 0.031 mmol) in ethyl acetate (3.0 ml), 20% Pd(OH)<sub>2</sub> on carbon (73.6 mg) was added, stirred for 15 h under hydrogen at atmospheric pressure at room temperature, and then, filtered through Celite. The filtrate was concentrated in vacuo, and separated by preparative thin layer chromatography (CHCl<sub>3</sub>–MeOH, 5:1). The part containing a product was eluted with CHCl<sub>3</sub>-MeOH (5:1) to give a mixture of 24a contaminated with silica gel. The mixture was dissolved in CHCl<sub>3</sub>-MeOH-0.1 M HCl aq. (5:10:4) to remove the silica gel, and extracted with CHCl<sub>3</sub>. The chloroform layer was concentrated in vacuo to give 24a (33.5 mg, 57%) as an amorphous film. IR:  $\nu_{\text{max}}$  (KBr) 3372, 1736, 1664, 1490, 1189 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD-CDCl<sub>3</sub>(10:1)) 7.41-7.32 (4H, m), 7.26-7.17 (6H, m), 5.44 (1H, dd, J=10.3, 9.5 Hz), 5.24 (1H, dd, J=10.3, 9.5 Hz), 5.27-5.18 (2H, m), 4.91 (1H, d, J=8.1 Hz), 4.73 (1H, q, J=8.8 Hz), 4.25 (1H, dd, J=5.9, 5.1 Hz), 4.15 (1H, d, J=5.9 Hz), 4.06 (1H, d, J=11.0 Hz), 3.68-3.58 (2H, m), 3.45 (1H, t, J=9.5 Hz), 2.59-2.40 (6H, m), 2.38-2.22 (4H, m), 2.14 (2H, t, J=7.3 Hz), 1.68-1.20 (120H, m), 0.90 (18H, t, J=7.3-6.6 Hz). High Resolution MS (FAB, positive), calcd for  $(M+Na)^{+}$ : C107H185N2O23PNa 1920.3003; found: 1920.2963.

2,6-Anhydro-3-deoxy-7-O-[2-deoxy-4-O-diphenylphosphono-2-[(R)-3-(dodecanoyloxy)tetradecanamido]-6-Omethyl-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-β-**D-glucopyranosyl]-3-**[(*R*)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-idoheptonic acid (24b). Compound 23b (29.0 mg, 0.013 mmol) was treated as described in the formation of 24a from 23a to give 24b (12.4 mg, 57%) as a powder. IR:  $\nu_{\rm max}$  (KBr) 3350 (broad), 1736 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) 7.41-7.36 (4H, m), 7.27-7.18 (6H, m), 5.44 (1H, t, J=9.5 Hz), 5.27-5.18 (2H, m), 4.95 (1H, d, J=8.8 Hz), 4.71 (1H, q, J=9.2 Hz), 4.27-4.22 (1H, m), 4.14 (1H, d, J=5.1 Hz), 4.04 (1H, d, J=11.0 Hz), 4.02-3.94 (1H, m), 3.93-3.78 (3H, m), 3.77-3.68 (2H, m), 3.53-3.44 (3H, m), 3.21 (3H, s), 2.61-2.40 (6H, m), 2.38-2.21 (4H, m), 2.13 (2H, t, J=7.3 Hz), 1.68–1.17 (120H, m), 0.90 (18H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $(M+Na)^{+}$ : 1934.3160:  $C_{108}H_{187}N_2O_{23}PNa$ found: 1934.3162. Anal. Calcd for C<sub>108</sub>H<sub>187</sub>N<sub>2</sub>O<sub>23</sub>P+2H<sub>2</sub>O: C, 66.55; H, 9.88; N, 1.44; P, 1.59. Found: C, 66.22; H, 9.63; N, 1.47; P, 1.62.

2,6-Anhydro-3-deoxy-7-*O*-[2,6-deoxy-4-*O*-diphenylphosphono-2-[(*R*)-3-(dodecanoyloxy)tetradecanamido]-6fluoro-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-[(*R*)-3-(hydroxy)tetradecanamido]-

4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-idoheptonic acid (24c). To a solution of 23c (16.5 mg, 0.007 mmol) in ethanol (2.0 ml), 20%  $Pd(OH)_2$  on carbon (19.0 mg) was added, stirred for 20 h under hydrogen at atmospheric pressure at room temperature, and then, filtered through Celite. The filtrate was concentrated in vacuo, and separated by preparative thin layer chromatography (CHCl<sub>3</sub>-MeOH, 8:1). The part containing a product was eluted with CHCl<sub>3</sub>-MeOH (5:1) to give a mixture of 24c contaminated with silica gel. The mixture was dissolved in CHCl<sub>3</sub>–MeOH–0.1 M HCl aq. (5:10:4) to remove the silica gel, and extracted with CHCl<sub>3</sub>. The chloroform layer was concentrated in vacuo to give 24c (8.1 mg, 58%) as an amorphous film. IR:  $\nu_{\text{max}}$  (KBr) 1736, 1661, 1189 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD–CDCl<sub>3</sub>(10:1)) 7.42–7.35 (4H, m), 7.26–7.16 (6H, m), 5.48 (1H, t, J=9.9 Hz), 5.29–5.17 (3H, m), 5.03 (1H, d, J=8.1 Hz), 4.72 (1H, t, J=9.2 Hz), 4.59–4.47 (2H, m), 4.42–4.33 (1H, m), 4.33– 4.23 (1H, m), 4.22–4.12 (1H, m), 4.07 (1H, d, J=11.7 Hz), 4.03-3.96 (1H, m), 3.95-3.78 (3H, m), 3.77-3.70 (1H, m), 3.43 (1H, t, J=9.9 Hz), 2.61-2.42 (6H, m), 2.36-2.25 (4H, m), 2.18–2.12 (2H, m), 1.68–1.20 (120H, m), 0.90 (18H, t, J=7.3-5.9 Hz). High Resolution MS (FAB, positive), calcd for  $C_{107}H_{184}N_2O_{22}FPNa$  (M+Na)<sup>+</sup>: 1922.2960; found: 1922.2898.

2,6-Anhydro-3-deoxy-7-O-[2-deoxy-2-[(R)-3-(dodecanoyloxy)tetradecanamido]-4-O-phosphono-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (4a). To a solution of **24a** (5.2 mg, 0.0027 mmol) in THF (1.0 ml), PtO<sub>2</sub> (6.3 mg) was added, stirred for 18 h under hydrogen at atmospheric pressure at 50°C, and then filtered. The filtrate was concentrated in vacuo, and separated by preparative thin layer chromatography (CHCl<sub>3</sub>-EtOH-AcOH-H<sub>2</sub>O, 8:5:1:1). The part containing a product was eluted with CHCl<sub>3</sub>-EtOH-AcOH-H<sub>2</sub>O (8:5:1:1) to give a mixture of 4a contaminated with silica gel. The mixture was dissolved in CHCl<sub>3</sub>-MeOH-0.1 M HCl aq. (5:10:4) to remove the silica gel, and extracted with CHCl<sub>3</sub>. The chloroform layer was concentrated in vacuo to give 4a (4.2 mg, 88%) as a powder.  $[\alpha]_D^{26} = -2.6$  (c=0.18, CHCl<sub>3</sub>). IR:  $\nu_{max}$  (KBr) 3351, 1734, 1662, 1468 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD-CDCl<sub>3</sub> (10:1)) 5.32-5.12 (4H, m), 4.68 (1H, d, J=8.8 Hz), 4.50 (1H, d, J=5.9 Hz), 4.33 (1H, dd, J=10.2, 5.1 Hz), 4.21 (1H, q, J=9.5 Hz), 4.08 (1H, d, J=10.3 Hz), 4.02-3.95 (1H, m), 3.90-3.77 (6H, m), 3.65-3.57 (1H, m), 3.50-3.45 (1H, m), 2.68-2.65 (2H, m), 2.53-2.40 (4H, m), 2.37-2.26 (6H, m), 1.68-1.24 (120H, m), 0.90 (18H, t, J=6.6 Hz). High Resolution MS (FAB, negative), calcd for  $C_{95}H_{176}N_2O_{23}P(M-H)^-$ : 1744.2402; found: 1744.2352.

**2,6-Anhydro-3-deoxy-7***-O***-[2-deoxy-2-[(***R***)-3-(dodecanoyloxy)tetradecanamido]-6-***O***-methyl-4-***O***-phosphono-3-***O***-[(***R***)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-[(***R***)-3-(hydroxy)tetradecanamido]-4-***O***-[(***R***)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (4b). Compound 24b (17.3 mg, 0.009 mmol) was treated as described in the formation of 4a** from 24a to give 4b (7.8 mg, 49%) as a powder.  $[\alpha]_D^{26}$ =-4.0 (c=0.20, CHCl<sub>3</sub>). IR:  $\nu_{max}$  (KBr) 3351 (broad), 1734, 1664 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD-CDCl<sub>3</sub> (5:1)) 5.30-5.20 (4H, m), 4.67

7701

(1H, d, J=8.8 Hz), 4.48 (1H, d, J=5.1 Hz), 4.37–4.33 (1H, m), 4.14–4.05 (2H, m), 4.01–3.96 (1H, m), 3.92–3.87 (2H, m), 3.82–3.76 (3H, m), 3.64–3.55 (3H, m), 3.42 (3H, s), 2.78–2.21 (12H, m), 1.67–1.23 (120H, m), 0.90 (18H, t, J=6.6 Hz). High Resolution MS (FAB, negative), calcd for C<sub>96</sub>H<sub>178</sub>N<sub>2</sub>O<sub>23</sub>P (M–H)<sup>-</sup>: 1758.2558; found: 1758.2534.

2,6-Anhydro-3-deoxy-7-O-[2,6-deoxy-2-[(R)-3-(dodecanoyloxy)tetradecanamido]-6-fluoro-4-O-phosphono-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (4c). Compound 24c (7.7 mg, 0.0040 mmol) was treated as described in the formation of 4a from 24a to give 4c (5.2 mg, 73%) as a powder.  $[\alpha]_D^{25} = -3.7$  (c=0.20, CHCl<sub>3</sub>). IR:  $\nu_{\text{max}}$  (KBr) 3362, 1734, 1662, 1467 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD-CDCl<sub>3</sub> (10:1)) 5.32-5.24 (4H, m), 4.68 (1H, d, J=9.5 Hz), 4.50 (1H, d, J=5.9 Hz), 4.33 (1H, dd, J=10.6, 6.2 Hz), 4.22 (1H, q, J=8.8 Hz), 4.07 (1H, d, J=9.5 Hz), 4.00-3.97 (1H, m), 3.90-3.85 (1H, m), 3.82-3.62 (6H, m), 3.60–3.48 (2H, m, including 1H, t, J=8.8 Hz at 3.60 ppm), 2.72–2.62 (2H, m), 2.55–2.40 (4H, m), 2.37– 2.23 (6H, m), 1.67–1.25 (120H, m), 0.90 (18H, t, J= 6.6 Hz). High Resolution MS (FAB, negative), calcd for  $C_{95}H_{175}N_2O_{22}FP (M-H)^-$ : 1746.2358; found: 1746.2345.

Diphenylmethyl 2,6-anhydro-7-O-[2-acetamido-6-O-benzyloxycarbonyl-2-deoxy-4-O-diphenylphosphono-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3-deoxy-D-glycero-D-ido-heptonate (25a). To a solution of 22a (99.9 mg, 0.046 mmol) in acetic acid (2.0 ml), zinc dust (310 mg, 4.74 mmol) was added. After vigorously stirring for 5 h at room temperature, the solution was filtered through Celite to remove the zinc dust and concentrated in vacuo to give a crude product. To a solution of this crude product in  $CH_2Cl_2$  (2.0 ml), acetic acid (12 mg) in  $CH_2Cl_2$  (2.0 ml) and EDCI·HCl (17.8 mg, 0.093 mmol) were added at room temperature, and the mixture was allowed to stand for 4 h at room temperature. Then, the reaction mixture was concentrated in vacuo, and chromatographed on a silica gel column eluted with hexane-EtOAc (2:1) to give 25a (43.5 mg, 47%) as a viscous oil. IR:  $\nu_{\rm max}$  (KBr) 3360, 1740, 1652, 1531, 1192 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) 7.35-7.11 (35H, m), 6.82 (1H, s), 6.53 (1H, d, J=3.9 Hz), 6.52 (1H, d, J=6.8 Hz), 5.50 (1H, t, J=9.3 Hz), 5.36-5.30 (2H, m), 5.18-5.08 (1H, m), 5.12 (1H, AB-q, J=12.2 Hz), 5.03 (1H, AB-q, J=12.2 Hz), 4.68-4.62 (2H, m, including 1H, AB-q, J=5.9 Hz at 4.62 ppm), 4.54 (1H, AB-q, J=11.7 Hz), 4.49-4.45 (2H, m, including 1H, AB-q, J=10.7 Hz at 4.47 ppm), 4.40-4.33 (3H, m), 4.18 (1H, dd, J=12.2, 4.4 Hz), 4.03 (2H, d, J=9.8 Hz), 3.88-3.81 (1H, m), 3.78-3.73 (1H, m), 3.68-3.55 (4H, m), 3.38-3.31 (2H, m), 2.61 (1H, dd, J=15.4, 7.3 Hz), 2.46-2.38 (2H, m), 2.28-2.11 (5H, m), 1.87 (3H, s), 1.75–1.08 (85H, m), 0.88 (12H, t, J=6.9 Hz). High Resolution MS (FAB, positive), calcd for C<sub>118</sub>H<sub>167</sub>N<sub>2</sub>O<sub>23</sub>PNa (M+Na)<sup>+</sup>: 2034.1595; found: 2034.1606. Anal. Calcd for C<sub>118</sub>H<sub>167</sub>N<sub>2</sub>O<sub>23</sub>P: C, 70.42; H, 8.36; N, 1.39; P, 1.54. Found: C, 70.14; H, 8.22; N, 1.28; P, 1.27.

# Diphenylmethyl 2,6-anhydro-7-*O*-[2-acetamido-2-deoxy-4-*O*-diphenylphosphono-6-*O*-methyl-3-*O*-[(*R*)-3-(tetra-

decanoyloxy)tetradecanoyl]-B-D-glucopyranosyl]-3-[(R)-3-(benzyloxy)tetradecanamido]-4-O-[(R)-3-(benzyloxy)tetradecanoyl]-3-deoxy-D-glycero-D-ido-heptonate (25b). Compound 22b (23.3 mg, 0.012 mmol) was treated as described in the formation of 25a from 22a to give 25b (15.9 mg, 75%) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3339, 1742, 1654 cm<sup>-1</sup>. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>) 7.39–7.13 (30H, m), 6.82 (1H, s), 6.52 (1H, d, J=9.0 Hz), 6.41 (1H, d, J=7.1 Hz), 5.46 (1H, dd, J=10.0, 9.4 Hz), 5.30 (1H, dd, J=10.5, 9.3 Hz), 5.22 (1H, d, J=8.2 Hz), 5.19-5.10 (1H, m), 4.72–4.40 (6H, m), 4.36 (2H, d, J=1.9 Hz), 4.05–3.58 (6H, m), 3.52-3.35 (4H, m), 3.21 (3H, s), 2.65-2.14 (8H, m), 1.87 (3H, s), 1.78-1.11 (82H, m), 0.88 (12H, t, J= 6.4 Hz). High Resolution MS (FAB, positive), calcd for  $C_{111}H_{163}N_2O_{21}PNa$  $(M+Na)^{+}$ : 1914.1384; found: 1914.1450. Anal. Calcd for C<sub>111</sub>H<sub>163</sub>N<sub>2</sub>O<sub>21</sub>P: C, 70.45; H, 8.68; N, 1.48; P, 1.64. Found: C, 70.50; H, 8.86; N, 1.45; P, 1.75.

Diphenylmethyl 2,6-anhydro-7-O-[2-acetamido-2,6-deoxy-4-O-diphenylphosphono-6-fluoro-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]- $\beta$ -D-glucopyranosyl]-3-[(R)-3-(benzyloxy)tetradecanamido]-4-*O*-[(*R*)-3-(benzyloxy)tetradecanoyl]-3-deoxy-D-glycero-D-ido-heptonate (25c). Compound 22c (122 mg, 0.061 mmol) was treated as described in the formation of 25a from 22a to give 25c (65.1 mg, 57%) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3356, 1733, 1654, 1533, 1193 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 7.36– 7.08 (30H, m), 6.82 (1H, s), 6.55 (1H, d, J=7.3 Hz), 6.52 (1H, d, J=9.5 Hz), 5.53 (1H, t, J=9.5 Hz), 5.38-5.28 (2H, m), 5.15–5.13 (1H, m), 4.67–4.60 (2H, m), 4.57–4.45 (4H, m), 4.39-4.32 (2H, m), 4.07-4.04 (1H, m), 3.86-3.83 (1H, m), 3.73-3.56 (5H, m), 3.40-3.33 (2H, m), 2.61 (1H, dd, J=7.3, 15.4 Hz), 2.41 (2H, dd, J=5.1, 15.4 Hz), 2.25-2.11 (5H, m), 1.87 (3H, s), 1.66-1.25 (82H, m), 0.88 (12H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{110}H_{160}N_2O_{20}FPNa$  (M+Na)<sup>+</sup>: 1902.1184; found: 1902.1167. Anal. Calcd for  $C_{110}H_{160}N_2O_{20}FP$ : C, 70.26; H, 8.58; N, 1.49; F, 1.01; P, 1.65. Found: C, 69.98; H, 8.08; N, 1.34; F, 1.12; P, 1.77.

2,6-Anhydro-7-O-[2-acetamido-2-deoxy-4-O-diphenylphosphono-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]- $\beta$ -D-glucopyranosyl]-3-deoxy-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-Dglycero-D-ido-heptonic acid (26a). To a solution of 25a (15.1 mg, 0.008 mmol) in ethanol (1.0 ml), 20% Pd(OH)<sub>2</sub> on carbon (16.0 mg) was added, and the mixture was stirred for 7 h under hydrogen at atmospheric pressure at room temperature, and filtered through Celite. The filtrate was concentrated in vacuo, and separated by preparative thin layer chromatography (CHCl<sub>3</sub>-MeOH, 5:1). The part containing a product was eluted with CHCl<sub>3</sub>-MeOH (5:1) to give a mixture of **26a** contaminated with silica gel. The mixture was dissolved in CHCl<sub>3</sub>-MeOH-0.1 M HCl aq. (5:10:4) to remove the silica gel, and extracted with CHCl<sub>3</sub>. The chloroform layer was concentrated in vacuo to give 26a (7.8 mg, 63%) as a powder. IR:  $\nu_{\text{max}}$  (KBr) 3373, 1738, 1662, 1490, 1189 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) 7.41-7.36 (4H, m), 7.26-7.18 (6H, m), 5.41 (1H, t, J=9.9 Hz), 5.30 (1H, t, J=10.3 Hz), 5.11 (1H, t, J=6.2 Hz), 4.76 (1H, d, J=8.8 Hz), 4.71 (1H, q, J=9.2 Hz), 4.49-4.46 (1H, m), 4.36-4.28 (1H, m), 4.11

(1H, d, J=11.0 Hz), 3.97–3.75 (6H, m), 3.67–3.58 (2H, m), 3.53 (1H, t, J=8.8 Hz), 2.54–2.39 (4H, m), 2.32–2.21 (2H, m), 2.16 (2H, t, J=7.3 Hz), 1.94 (3H, s), 1.53–1.18 (82H, m), 0.91–0.88 (12H, m). High Resolution MS (FAB, positive), calcd for  $C_{83}H_{139}N_2O_{21}PNa$  (M+Na)<sup>+</sup>: 1553.9506; found: 1553.9490. Anal. Calcd for  $C_{83}H_{139}N_2O_{21}P+2H_2O$ : C, 63.58; H, 9.19; N, 1.79; P, 1.98. Found: C, 63.21; H, 8.72; N, 1.77; P, 2.02.

2,6-Anhydro-7-O-[2-acetamido-2-deoxy-4-O-diphenylphosphono-6-O-methyl-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-deoxy-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (26b). To a solution of 25b (14.6 mg, 0.008 mmol) in EtOAc (3.0 ml), 20% Pd(OH)<sub>2</sub> on carbon (15.2 mg) was added, and the mixture was stirred for 16 h under hydrogen at atmospheric pressure at room temperature, and then, filtered through Celite. The filtrate was concentrated in vacuo, and separated by preparative thin layer chromatography (CHCl<sub>3</sub>–MeOH, 8:1). The part containing a product was eluted with CHCl<sub>3</sub>-MeOH (5:1) to give a mixture of **26b** contaminated with silica gel. The mixture was dissolved in CHCl<sub>3</sub>-MeOH-0.1 M HCl aq. (5:10:4) to remove the silica gel, and extracted with CHCl<sub>3</sub>. The chloroform layer was concentrated in vacuo to give **26b** (7.7 mg, 63%) as a powder. IR:  $\nu_{max}$  (KBr) 3500-3300 (broad), 1740,  $1655 \text{ cm}^{-1}$ . <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) 8.16 (1H, d, J=10.0 Hz), 7.41-7.37 (4H, m), 7.27-7.19 (6H, m), 5.41 (1H, t, J=9.9 Hz), 5.26 (1H, t, J=9.5 Hz), 5.10 (1H, t, J=5.9 Hz), 4.97 (1H, d, J=8.1 Hz), 4.72 (1H, q, J=9.2 Hz), 4.27-4.23 (1H, m), 4.16 (1H, d, J=5.1 Hz), 4.04 (1H, d, J=12.5 Hz), 3.97–3.82 (4H, m), 3.77-3.70 (2H, m), 3.54-3.41 (3H, m), 3.22 (3H, s), 2.50-2.37 (4H, m), 2.34–2.21 (2H, m), 2.15 (2H, t, J=7.3 Hz), 2.00 (3H, s), 1.55–1.25 (82H, m), 0.90 (12H, t, J=7.3–6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{84}H_{141}N_2O_{21}PNa (M+Na)^+$ : 1567.9662; found: 1567.9694. Anal. Calcd for C<sub>84</sub>H<sub>141</sub>N<sub>2</sub>O<sub>21</sub>P: C, 65.26; H, 9.19; N, 1.81; P, 2.00. Found: C, 64.89; H, 9.04; N, 1.64; P, 1.97.

2,6-Anhydro-7-O-[2-acetamido-2,6-deoxy-4-O-diphenylphosphono-6-fluoro-3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]- $\beta$ -D-glucopyranosyl]-3-deoxy-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (26c). Compound 25c (54.5 mg, 0.029 mmol) was treated as described in the formation of 26a from 25a to give 26c (29.1 mg, 65%) as an amorphous film. IR:  $\nu_{\rm max}$  (KBr) 3345, 1738, 1662, 1491, 1189 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) 7.41-7.37 (4H, m), 7.27-7.17 (6H, m), 5.46 (1H, t, J=9.9 Hz), 5.27 (1H, t, J=9.5 Hz), 5.13-5.04 (1H, m), 4.81 (1H, d?, unclear due to overlapping with solvent's peak), 4.70 (1H, q, J=9.3 Hz), 4.60 (1H, d, J=11.0 Hz), 4.49 (1H, d, J=11.0 Hz), 4.40–4.36 (1H, m), 4.27–4.23 (1H, m), 4.18–4.14 (1H, m), 4.04 (1H, d, J=12.5 Hz), 3.99-3.81 (4H, m), 3.76-3.68 (1H, m), 3.42 (3H, t, J=9.5 Hz), 2.50-2.41 (4H, m), 2.34-2.21 (2H, m), 2.16 (2H, t, J=7.3 Hz), 2.01 (3H, s), 1.55–1.20 (82H, m), 0.90 (12H, t, J=6.6 Hz). High Resolution MS (FAB, positive), calcd for  $C_{83}H_{138}N_2O_{20}FPNa (M+Na)^+$ : 1555.9462; found: 1555.9437. Anal. Calcd for C<sub>83</sub>H<sub>138</sub>N<sub>2</sub>O<sub>20</sub>FP+2H<sub>2</sub>O: C, 63.50; H, 9.12; N, 1.78; F, 1.21; P, 1.97. Found: C, 63.37; H, 8.66; N, 1.76; F, 1.41; P, 2.11.

2,6-Anhydro-7-O-[2-acetamido-2-deoxy-4-O-phosphono- $3-O-[(R)-3-(tetradecanoyloxy)tetradecanoyl]-\beta-D-gluco$ pyranosyl]-3-deoxy-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hvdroxy)tetradecanovl]-D-glycero-Dido-heptonic acid (5a). To a solution of 26a (6.6 mg, 0.004 mmol) in THF (1.0 ml), PtO<sub>2</sub> (8.4 mg) was added, and the mixture was stirred for 20 h under hydrogen at atmospheric pressure at room temperature, and then filtered. The filtrate was concentrated in vacuo, and separated by preparative thin layer chromatography (CHCl<sub>3</sub>-EtOH-AcOH-H<sub>2</sub>O, 8:5:1:1). The part containing a product was eluted with CHCl<sub>3</sub>-EtOH-AcOH-H<sub>2</sub>O (8:5:1:1) to give a mixture of 5a contaminated with silica gel. The mixture was dissolved in CHCl<sub>3</sub>-MeOH-0.1 M HCl aq. (5:10:4) to remove the silica gel, and extracted with CHCl<sub>3</sub>. The chloroform layer was concentrated in vacuo to give 5a (4.9 mg, 82%) as a powder.  $[\alpha]_{D}^{25} = -7.8$  (c=0.20, CHCl<sub>3</sub>). IR:  $\nu_{\text{max}}$  (KBr) 3368, 1737, 1664, 1466 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD-CDCl<sub>3</sub> (10:1)) 5.26-5.12 (3H, m), 4.62 (1H, d, J=8.8 Hz), 4.31-4.18 (3H, m), 4.09 (1H, d, J=9.5 Hz), 4.03-3.87 (4H, m), 3.82-3.73 (3H, m), 3.50-3.38 (2H, m), 2.74–2.60 (2H, m), 2.52–2.38 (2H, m), 2.36– 2.22 (4H, m), 1.98 (3H, s), 1.63-1.58 (4H, m), 1.45-1.41 (6H, m), 1.38–1.23 (72H, m), 0.90 (12H, t, *J*=7.3–6.6 Hz). High Resolution MS (FAB, negative), calcd for  $C_{71}H_{130}N_2O_{21}P$  (M–H)<sup>-</sup>: 1377.8904; found: 1377.8844. Anal. Calcd for C<sub>71</sub>H<sub>131</sub>N<sub>2</sub>O<sub>21</sub>P+H<sub>2</sub>O: C, 61.01; H, 9.59; N, 2.00; P, 2.22. Found: C, 61.24; H, 9.79; N, 2.14; P, 1.94.

2,6-Anhydro-7-O-[2-acetamido-2-deoxy-6-O-methyl-4-*O*-phosphono-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-deoxy-3-[(R)-3-(hydroxy)tetradecanamido]-4-O-[(R)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (5b). Compound 26b (7.0 mg, 0.0045 mmol) was treated as described in the formation of 5a from 26a to give 5b (6.8 mg, 99%) as a powder.  $[\alpha]_{D}^{26} = -6.6$  (c=0.20, CHCl<sub>3</sub>). IR:  $\nu_{max}$  (KBr) 3362, 1737, 1665 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD) 5.30 (1H, dd, J=10.7, 8.8 Hz), 5.23–5.15 (2H, m), 4.66 (1H, d, J=8.5 Hz), 4.50 (1H, d, J=5.7 Hz), 4.34-4.29(1H, m), 4.22–4.19 (1H, m), 4.10 (1H, d, J=1.4 Hz), 4.00-3.94 (1H, m), 3.88-3.73 (5H, m), 3.62-3.57 (2H, m), 3.54 (1H, t, J=9.1 Hz), 3.41 (3H, s), 2.72-2.22 (8H, m), 1.94 (3H, s), 1.62-1.56 (4H, m), 1.48-1.40 (6H, m), 1.39–1.23 (72H, m), 0.91 (12H, t, J=7.0–6.6 Hz). High Resolution MS (FAB, negative), calcd for C<sub>72</sub>H<sub>132</sub>N<sub>2</sub>O<sub>21</sub>P (M-H)<sup>-</sup>: 1391.9060; found: 1391.9093. Anal. Calcd for C<sub>72</sub>H<sub>133</sub>N<sub>2</sub>O<sub>21</sub>P+H<sub>2</sub>O: C, 61.25; H, 9.64; N, 1.98; P, 2.19. Found: C, 61.53; H, 9.50; N, 1.99; P, 2.23.

**2,6-Anhydro-7-***O*-[**2**-acetamido-2,6-deoxy-6-fluoro-4-*O*phosphono-3-*O*-[(*R*)-3-(tetradecanoyloxy)tetradecanoyl]-β-D-glucopyranosyl]-3-deoxy-3-[(*R*)-3-(hydroxy)tetradecanamido]-4-*O*-[(*R*)-3-(hydroxy)tetradecanoyl]-D-glycero-D-ido-heptonic acid (5c). Compound **26c** (17.1 mg, 0.0111 mmol) was treated as described in the formation of **5a** from **26a** to give **5c** (13.3 mg, 87%) as a powder.  $[\alpha]_D^{26}$ =-18.6 (c=0.20, CHCl<sub>3</sub>). IR:  $\nu_{max}$  (KBr) 3363, 1734, 1662, 1466 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD-CDCl<sub>3</sub>(10:1)) 5.23-5.14 (3H, m), 4.72-4.57 (2H, m), 4.33-4.28 (2H, m), 4.18-4.12 (2H, m), 4.00-3.88 (3H, m), 3.78-3.56 (4H, m), 3.32 (1H, t, J=1.5 Hz), 2.74 (1H, d, J=9.5, 6.6 Hz), 2.63 (1H, dd, J=9.5, 6.6 Hz), 2.52-2.40 (2H, m), 2.36–2.22 (4H, m), 1.99 (3H, s), 1.63–1.58 (4H, m), 1.50–1.42 (6H, m), 1.38–1.22 (72H, m), 0.89 (12H, t, J=7.3-6.6 Hz). High Resolution MS (FAB, negative), calcd for C<sub>71</sub>H<sub>129</sub>N<sub>2</sub>O<sub>20</sub>FP (M–H)<sup>-</sup>: 1379.8860; found: 1379.8854. Anal. Calcd for C<sub>71</sub>H<sub>130</sub>N<sub>2</sub>O<sub>20</sub>FP: C, 61.72; H, 9.48; N, 2.03; F, 1.38; P, 2.24. Found: C, 61.55; H, 9.36; N, 1.76; F, 1.19; P, 2.11.

## Method for biological activity measurement<sup>12</sup>

The sources of the materials used in the study are as follows: lipopolysaccharide (LPS) from *E. coli* serotype 026:B6<sup>18</sup> and 12-*O*-tetradecanoylphorbor acetate (TPA) were from SIGMA, St. Louis, MO; RPMI-1640 medium, fetal bovine serum (FBS), and newborn calf serum (NBCS) were from GIBCO, Grand Island, NY; and human tumor necrosis factor- $\alpha$  enzyme-linked immunosorbent assay (TNF $\alpha$ ELISA) kit was from Genzyme, Cambridge, MA.

Cell culture: Human monoblastic U937 cells were maintained in RPMI-1640 medium supplemented with 10% FBS, 100 units/ml of penicillin and 100 µg/ml of streptomycin (growth medium).

Production of TNF $\alpha$  by U937 cells: U937 cells (1×10<sup>4</sup>/ 200µl/well) were plated in 96-well plates (Corning, Cambridge, MA), and were cultured in the presence of TPA (30 ng/ml) for 72 h at 37°C. After removing the supernatant, the cells were incubated in 200 µl of fresh RPMI-1640 medium containing 10% NBCS, in the absence or the presence of 30 ng/ml of LPS with graded concentrations of the compounds in a humidified atmosphere of 5%  $CO_2$  for 4.5 h at 37°C. After incubation, the amount of TNF $\alpha$ produced in the culture supernatants was determined using the TNF $\alpha$  ELISA kits. As a control, the amount of TNF $\alpha$ produced by the U937 cells, which were stimulated with 30 ng/ml of LPS in the absence of compounds, was used. The concentrations (nM) of compounds required to inhibit the LPS-induced TNF $\alpha$  production by U937 cells by 50%  $(IC_{50})$  was calculated from the control amount. All experiments were carried out at least twice, showing the data are reproducible. The representative data are shown in Figs. 3 and 4.

#### Acknowledgements

This study was supported, in part, by the Social Insurance Agency Fund commissioned by the Japan Health Sciences Foundation.

#### References

 Westphal, O.; Lüderitz, O. Angew. Chem. 1954, 66, 407–417.
 Galanos, C.; Lüderitz, O.; Rietschel, E. T.; Westphal, O. Int. Rev. Biochem. 1977, 14, 239–335. 3. (a) Imoto, M.; Yoshimura, H.; Sakaguchi, N.; Kusumoto, S.; Shiba, T. *Tetrahedron Lett.* **1985**, *26*, 1545–1548. (b) Imoto, M.; Yoshimura, H.; Shimamoto, T.; Sakaguchi, N.; Kusumoto, S.; Shiba, T. *Bull. Chem. Soc. Jpn* **1987**, *60*, 2205–2214.
(c) Takahashi, T.; Nakamoto, S.; Ikeda, K.; Achiwa, K. *Tetrahedron Lett.* **1986**, *27*, 1819–1822.

4. Galanos, C. Z. Immunitatsforsch. Exp. Klin. Immunol. 1975, 149, 214–229.

Rietschel, E. T.; Kirikae, T.; Schade, F. U.; Ulmer, A. J.; Holst,
 O.; Brade, H.; Schmidt, G.; Mamat, U.; Grimmecke, H-D.;
 Kusumoto, S.; Zähringer, U. *Immunobiology* 1993, *187*, 169–190.
 (a) Qureshi, N.; Honovich, J. P.; Hara, H.; Cotter, R. J.;
 Takayama, K. *J. Biol. Chem.* 1988, *263*, 5502–5504. (b) Qureshi,
 N.; Takayama, K.; Kurtz, R. *Infect. Immun.* 1991, *59*, 441–444.

7. (a) Christ, W. J.; McGuinness, P. D.; Asano, O.; Wang, Y.; Mullarkey, M. A.; Perez, M.; Hawkins, L. D.; Blythe, T. A.; Dubuc, G. R.; Robidoux, A. L. *J. Am. Chem. Soc.* **1994**, *116*, 3637–3638. (b) Christ, W. J.; Asano, O.; Robidoux, A. L. C.; Perez, M.; Wang, Y.; Dubuc, G. R.; Gavin, W. E.; Hawkins, L. D.; McGuinness, P. D.; Mullarkey, M. A.; Lewis, M. D.; Kishi, Y.; Kawata, T.; Bristol, J. R.; Rose, J. R.; Rossignol, D. P.; Kobayashi, S.; Hishinuma, I.; Kimura, A.; Asakawa, N.; Katayama, K.; Yamatsu, I. *Science* **1995**, *268*, 80–83.

8. Kusama, T.; Soga, T.; Ono, Y.; Kumazawa, E.; Shioya, E.; Nakayama, K.; Uoto, K.; Osada, Y. *Chem. Pharm. Bull.* **1991**, *39*, 3244–3253.

9. Liu, W.-C.; Oikawa, M.; Fukase, K.; Suda, Y.; Kusumoto, S. *Bull. Chem. Soc. Jpn* **1999**, *72*, 1377–1385.

10. Wang, M.-H.; Flad, H-D.; Feist, W.; Musehold, J.; Kusumoto, S.; Brade, H.; Gerdes, J.; Rietschel, H. T.; Ulmer, A. J. *Lymphokine Cytokine Res.* **1992**, *11*, 23–31.

 (a) Matsuura, M.; Kojima, Y.; Homma, J. Y.; Kubota, Y.; Yamamoto, A.; Kiso, M.; Hasegawa, A. *FEBS Lett.* **1984**, *167*, 226–230. (b) Kiso, M.; Ogawa, Y.; Fujishima, Y.; Fujita, M.; Tanaka, S.; Hasegawa, A. *J. Carbohydr. Chem.* **1987**, *6*, 625–638.
 Shiozaki, M.; Kurakata, S.; Tatsuta, T.; Maeda, H.; Nishijima, M. *Tetrahedron* **1997**, *53*, 16041–16060.

13. Mochizuki, T.; Iwano, Y.; Shiozaki, M.; Kurakata, S.; Kanai, S.; Nishijima, M. *Carbohydr. Res.* **2000**, *324*, 225–230.

14. Shiozaki, M.; Deguchi, N.; Macindoe, W. M.; Arai, M.; Miyazaki, H.; Mochizuki, T.; Tatsuta, T.; Ogawa, J.; Maeda, H.; Kurakata, S. *Carbohydr. Res.* **1996**, *283*, 27–51.

15. (a) Haines, L. M.; Singleton, E. J. Chem. Soc., Dalton Trans.
1972, 1891–1896. (b) Oltvoort, J. J.; van Boeckel, C. A. A.; de Koning, J. H.; van Boom, J. H. Synthesis 1981, 305–308.

16. Schmidt, R. R. Angew. Chem. Int. Ed. Engl. 1986, 25, 212–235.

17. Shiozaki, M.; Arai, M.; Macindoe, W. M.; Mochizuki, T.; Wakabayashi, T.; Kurakata, S.; Tatsuta, T.; Maeda, H.; Nishijima, M. *Bull. Chem. Soc. Jpn* **1997**, *70*, 1149–1161.

18. (a) Dickerson, R.; Manzo, C.; Charland, S.; Settle, R.; Stein,

T.; Kuhl, D.; Rajter, J. *J. Surg. Res.* **1995**, *58*, 260–266. (b) Michel, O.; Duchateau, J.; Plat, G.; Cantinieaux, B.; Hotimsky, A.; Gerain, J.; Sergysels, R. *Clin. Exp. Allergy* **1995**, *25*, 73–79.